



Review Article

Epigenetically Regulating Non-coding RNAs in Colorectal Cancer: Promises and Potentials

Zahra Taheri¹ , Majid Zaki-Dizaji^{2*} ¹Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran²Human Genetics Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran**Abstract**

Colorectal cancer (CRC) is a common malignancy with high mortality. Despite advancements in understanding its molecular causes and improved drug therapies, patient survival rates remain low. The main reasons for the high mortality rate are cancer metastasis and the emergence of drug-resistant cancer cell populations. While genetic changes are recognized as the main driver of CRC occurrence and progression, recent studies suggest that epigenetic regulation is a crucial marker in cancer, influencing the interplay between genetics and the environment. Research has shown the significant regulatory roles of non-coding RNAs (ncRNAs) in CRC development. This review explores epigenetically regulated ncRNAs and their functions, aiming to understand key regulatory mechanisms that impact CRC development. Additionally, it discusses the potential use of these ncRNAs in CRC diagnosis, prognosis, and targeted treatments.

Keywords: Colorectal cancer, Non-coding RNA, Epigenetic, DNA methylation, Epitranscriptome, RNA modification

Cite this article as: Taheri Z, Zaki-Dizaji M. Epigenetically regulating non-coding RNAs in colorectal cancer: promises and potentials. *Middle East J Dig Dis* 2025;17(1):40-53. doi: 10.34172/mejdd.2025.404.

Received: July 27, 2024, **Accepted:** December 9, 2024, **ePublished:** January 31, 2025

Introduction

Colorectal cancer (CRC), also known as colon or rectal cancer, is a common and aggressive form of cancer. It ranks as the fourth leading cause of cancer-related death globally, following lung, liver, and stomach cancer.^{1,2} Most patients either have metastases upon diagnosis or develop them later due to the natural progression of the disease.³ Despite significant progress in radiotherapy, chemotherapy, and surgical procedures for CRC, as well as improvements in screening programs and medical technologies, the overall survival rate of patients with CRC remains relatively low. Thus, a clinical imperative exists to enhance our understanding of the biological processes in CRC that lead to gene deregulation, tumor heterogeneity, and evasion of drug treatment effects. It is essential to identify new disease determinants and utilize them as biomarkers for early disease detection, predicting drug responses, and prognosis.⁴

CRC is mostly a complex disease influenced by genetic and epigenetic risk factors and environmental factors that can impact its progression.⁵ Increasing evidence indicates that epigenetic changes play a role in shaping both normal physiological processes and the development of diseases, particularly in carcinogenesis.⁶ Epigenetic genome modifications are dynamic and reversible, including DNA, RNA, and chromatin modifications.⁶⁻⁸ We know that over 90% of the human genome is actively transcribed. Nevertheless, only 2% of these transcripts code for proteins, with the majority being non-coding

RNAs (ncRNAs).⁹ These ncRNAs include microRNA (miRNA), small nuclear RNA (snRNA), small nucleolar RNA (snoRNA), piwi-interacting RNA (piRNA), long non-coding RNAs (lncRNAs), ribosomal RNA (rRNA), transfer RNA (tRNA), and circular RNA (circRNA).¹⁰⁻¹³ Also, two new classes of ncRNAs, known as promoter-associated RNAs (PARs) and enhancer RNAs (eRNAs), have recently been identified.¹⁴ The impact of ncRNAs in CRC is well-known,¹⁵ and these ncRNAs can be epigenetically regulated during CRC development, metastasis, and drug.⁷

Because of the significance of epigenetic mutations (epimutations) in CRC and the increasing evidence of how epigenetically regulated ncRNAs contribute to CRC development, progression, and resistance, our objective is to outline recent discoveries, assess their molecular roles, and consider their potential as biomarkers for diagnosis, prognosis, and therapy.

Epigenetically Regulated MicroRNA Markers in CRC

MicroRNAs are short, ncRNAs, typically 20-25 nucleotides in length, that are crucial in biological processes such as regulating gene expression and various cellular processes like cell proliferation, cycle cell regulation, apoptosis, and differentiation.¹⁶⁻¹⁸ MiRNAs are present in all tissues, most binding to specific target mRNAs through the 3' UTR to downregulate gene expression or inhibit translation. However, reports suggest that miRNAs interact with various regions, such as promoters, the 5' UTR, and the



*Corresponding Author: Majid Zaki-Dizaji, Email: majid.zakidizaji@gmail.com



© 2025 The Author(s). This work is published by Middle East Journal of Digestive Diseases as an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

coding sequence.¹⁶ Additionally, microRNAs serve dual purposes as both oncogenes and tumor suppressors, playing a crucial role in the development of tumors.¹⁹ Dysregulation of miRNAs is linked to numerous human diseases.^{20,21} Epigenetic regulation, specifically DNA methylation, plays a crucial role in suppressing miRNAs. The abnormal methylation of miRNAs is a new type of biomarker that shows promise for diagnosing and prognostic markers in CRC. Recent studies have identified them as potential biomarkers for diagnosis and prognosis, as listed in Table 1 and Figure 1. Here, we discuss recently reported methylated miRNAs and their impact on CRC progression.

MiRNA-124

MiR-124 is an 85-base miRNA found on the minus strand of chromosome 8p23.1. It is predominantly expressed in the CNS and plays a crucial role in synaptic transmission, neuronal differentiation, stem cell regulation, and gastrulation.^{22,64,65} Researchers have found that miR-124 plays a tumor-suppressing role in different types of cancers, including colorectal and lung cancer. This is due to its specific methylation in tumors, low expression, hypomethylation in normal tissues, and high expression.^{22,66} Additionally, miR-124 is the initial microRNA in CRC that has been proven to be silenced through an epigenetic process.^{22,67} When the

Table 1. Epigenetically regulated miRNAs in colorectal cancer

miRNA Expression/ pattern in CRC	Target gene/ signaling pathway	Oncogene or Tumor- suppressor / biomarker	Sample type	Findings	Ref
miR-124a↓	CDK6	TS / Prognostic	cell lines and 208 CRC	The hypermethylation of miR-124a results in the activation of the CDK6 oncogene and phosphorylation of Rb.	2007 ²²
miR-342 ↓	-	TS	42 CRCs, 9 A, and 16 N	Methylated EVL/miR-342 was identified in a majority of CRCs, which suggests that it is an early event in CRC carcinogenesis.	2008 ²³
miR-34b/c ↓	MET, CCNE2, SFRS2 and CDK4	TS	111 CRC and cell lines	The CpG island of miR-34b/c acts as a bidirectional promoter controlling the expression of different tumor suppressor genes like BTG4. This region is often methylated in CRC.	2008 ²⁴
miR-345 ↓	BAG3	TS	CRC cells line	Low expression of mir-345 was associated with lymph node metastasis and worse histological type. Mir-345 acts as a growth inhibitor in CRC by targeting the BAG3 oncogene. This suggests its potential antineoplastic role in the development of CRC.	2011 ²⁵
miR-373 ↓	RAB22A	TS	CRC cell lines and 40 CRC	miR-373 downregulated and RAB22A upregulated in CRC	2011 ²⁶
miR-149 ↓	SP1	TS / Prognostic	86 CRC tissues and cell lines	Silencing miR-149 through methylation leads to the upregulation of Sp1, promoting CRC.	2013 ²⁷
miR-34a ↓	c-Met, Snail, β-catenin, CD44 ²⁸ in PC, AxI, TPD52, Lef1 and MTA2 ²⁹	TS / Prognostic	94 CRC w/o liver metastasis	Hypermethylation of miR-34a causes elevated levels of c-Met, Snail, and β-catenin, which are linked to liver metastasis in CRC.	2013 ³⁰
miR-497/195 ↓	miR-497: IGF1 ³¹ miR-195: BCL2 ³²	TS	CRC cell lines and 50 polyps with PANS	Both miRNAs are hypermethylated and expressed at lower levels in CRC. The tumor-suppressor activity of miR-497 in CRC is achieved by reducing the expression of IGF1. miR-195 targets BCL2, and the decreased expression of miR-195 has been strongly associated with higher mortality rates in CRC patients.	2013 ³³
miR-27↓	VEGFC	TS	CRC cell lines	miR-27b, found in CRC stem cells, functions as a crucial tumor suppressor and angiogenic factor by targeting VEGFC.	2013 ³⁴
miR-212 ↓	MnSOD	TS / Prognostic	180 CRC with PANS and cell lines	miR-212 inhibits metastasis and EMT in CRC by targeting MnSOD. The low level of miR-212 is linked to aggressive tumor behavior and a negative disease progression.	2013 ³⁵
miR-126 ↓	VEGF	TS / Therapeutic target	12 CRC with PANS 62 CRC and cell lines	miR-126 directly silences VEGF expression, leading to the inhibition of cell invasion and tumor angiogenesis in CRC.	2013 ³⁶
miR-638 ↓	TSPAN1, SOX2 ^{37,38} Sp2 ³⁹ in GC	TS / Prognostic	cell lines and 156 CRC with PANS	Downregulation of miR-638 in CRC was associated with poor prognoses. miR-638 inhibited CRC cell growth, invasion, and cell cycle progression by targeting TSPAN1.	2014 ⁴⁰
miR-204-5p↓	RAB22A	TS / Prognostic	CRC Cell lines and 272 CRC with PANS	Downregulation of miR-204-5p in CRC was associated with poor prognoses. miR-204-5p plays a role in inhibiting EMT.	2014 ⁴¹
miR-128 ↓	NEK2	TS / Prognostic	180 CRC and cell lines	MiR-128 inhibited NEK2 expression and cancer cell proliferation via cell cycle arrest. High miR-128 expression is associated with a low recurrence rate.	2014 ⁴²
miR-132 ↓	Paxillin, ZEB2 ⁴³	TS / Prognostic	36 CRC with PANS and cell lines	Downregulation of miR-132 may occur as a result of hypermethylation and implies a poor prognosis in CRC. miR-132 suppresses cell invasion and EMT in CRC by directly targeting ZEB2.	2015 ⁴⁴
miR-125a,b ↓	MUC1, ERBB2, and ERBB3 in BC ⁴⁵	TS / Prognostic	68 CRC with PANS	Hypermethylation of miR-125 was found to have a negative impact on the clinical outcome of patients.	2015 ⁴⁶
miR-133b↓	-	TS	CRC cell lines and CRC tissues	Methylation of miR-133b disrupts apoptosis, cell cycle progression, and invasion in CRC cells.	2015 ⁴⁷

Table 1. Continued.

miRNA Expression/ pattern in CRC	Target gene/ signaling pathway	Oncogene or Tumor- suppressor / biomarker	Sample type	Findings	Ref
miR-4500 ↓	HMGA2	TS / Prognostic	75 CRC with PANS	Downregulated miR-4500 indicated an advanced tumor stage and poor survival. In vitro, miR-4500 suppressed CRC cell proliferation, migration, and invasion, while in vivo, it inhibited tumor growth by targeting HMGA2.	2016 48
miR-23b ↓	PLAU, c-met ⁴⁹ in HCC SMADs/TGF- β/ BMP ⁵⁰ in liver stem cells	TS / Diagnostic and Prognostic	96 CRC plasma	Decreased levels of plasma miR-23b were strongly linked to clinical stage, tumor depth, distant metastasis, and tumor recurrence and ultimately shorter recurrence-free survival times and lower overall survival rates.	2016 51
miR-1247 ↓	MYCBP2	TS / Therapeutic target	35 CRC and cell line	miR-1247 suppresses tumor growth by targeting the oncogene MYCBP2 and its downstream c-myc in methylator CRC, effectively inhibiting tumor progression.	2018 52
miR-21-5P ↑	TET1 PTEN, PDCD4, SPRY2 ⁵³	Onco / Diagnostic and Prognostic	164 CRC and cell lines	TET1 acts as a suppressor of tumor growth and inhibits EMT. miR-21-5P directly targets TET1, leading to the promotion of EMT, migration, and invasion in CRC.	2018 54
miR-34c-5p ↓	SATB2	TS / Prognostic	85 CRC with PANS and cell lines	miR-34c-5p targets SATB2, reducing metastasis and inhibiting EMT in CRC.	2018 55
miR-137 ↓ miR-342 ↓	-	TS / Diagnostic and Prognostic	51 polyps, 8 CRC, and 14 N	The methylation frequency of miR-342 was higher than miR-137	2019 56
miR-200c/141 ↓	ZEB1, ZEB2 ⁵⁷	TS / Prognostic	34 CRC, 60 polyps with PANS, 20 N	DNA methylation of the miR-200c/141 cluster correlated with tumor stage and poor prognosis. The miR-200c-141 cluster is crucial in inhibiting EMT by targeting ZEB1 and ZEB2.	2019 58, 59
miR-152-3p ↓	TMSB10	TS / Prognostic	88 CRC	DNMT1 and TMSB10 upregulated and miR-152-3p downregulated in CRC. DNMT1 maintained methylation of miR-152-3p	2020 60
miR-34a ↓	CSF1R, PDGFR ^{61, 62}	TS	CRC cell lines	P53 suppresses CRC invasion, EMT, and metastasis partly through downregulation of CSF1R by inducing miR-34a. CSF1R is directly and indirectly induced by SNAIL.	2020 63

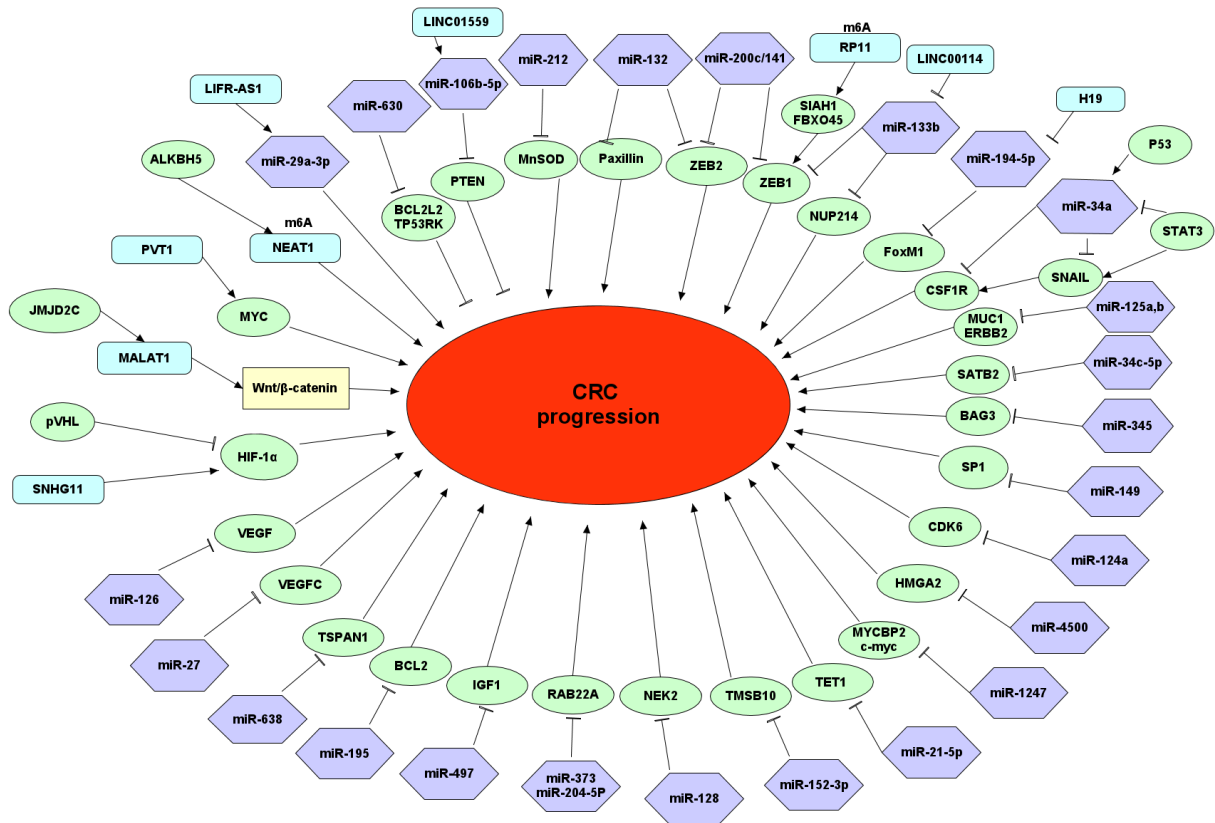


Figure 1. Impact of epigenetically regulated non-coding RNAs on colorectal cancer molecular pathways. The blue rectangles show lncRNAs, the green circles show genes, the purple hexagons are miRNAs, and the yellow rectangles are pathways. N6-methyladenosine (m6A)

epigenetic mechanism leads to hypermethylation of CpG island in promoter miR-124a, it leads to the activation

of an oncogene called CDK6 and the phosphorylation and suppression of a tumor suppressor gene known

as Rb. CDK6 is essential for cell cycle progression and differentiation. Its suppression through miR-124 could serve as a valuable biomarker for cancer prognosis and the development of anticancer treatments.²² Studies have shown that miR-124 methylation in bowel lavage fluid (BLF) is altered in patients with CRC, suggesting methylated miR-124-3 may be a potential non-invasive diagnostic biomarker for CRC.⁶⁸ Zhou and co-workers showed that reducing miR-124 levels could lead to increased cell growth, movement, invasion, and spread in CRC by inhibiting ROCK1 expression.⁶⁶ This evidence shows that miR-124 serves as a prognostic biomarker in patients with CRC.

MiR-126

MiR-126 is recognized as a key regulator of angiogenesis. New studies have uncovered conflicting roles of miR-126 in cancer development. Research has demonstrated that miR-126 functions as a tumor suppressor by reducing tumor cell growth and spreading by targeting oncogenes like ADAM9, SLC7A5, and Crk.³⁶ In contrast, recent studies suggest that miR-126 can play an oncogenic role by encouraging gastric carcinogenesis by suppressing SOX2 expression.⁶⁹ The luciferase reporter assay showed that miR-126 directly binds to the 3'UTR of VEGF mRNA, leading to the inhibition of cell migration, invasion, and tumor neovascularization caused by CRC cells.⁷⁰ Recent studies have indicated a reduction in miR-126 expression in CRC.⁷¹ Bioinformatic prediction tools have shown a conserved binding site for miR-126 in the 3'UTR of VEGF mRNA. The findings indicate that the silencing of miR-126 through promoter methylation is a significant factor in the disruption of VEGF expression in CRC. Therefore, miR-126 could potentially be a prognostic biomarker and targeted for CRC treatment.

MiR-1247

MiR-1247 is a novel miRNA that acts as a tumor suppressor and is found on the minus strand of 14q32.31. New research has found that MYC binding protein 2 (MYCBP2) is a gene targeted by miR-1247 in colon cancer. MYCBP2, is a protein that binds directly to the proto-oncogene c-myc and is involved in differentiation, cellular proliferation, and apoptosis. The specific molecular mechanisms triggered by MYCBP2 remain largely unidentified, but a significant finding is a substantial reduction in c-myc expression observed in cells with miR-1247 overexpression.^{52,72,73} Liang and colleagues conducted a study using tumor samples from patients with hypermethylated and non-methylated colon cancer, as well as cell lines. They discovered a correlation between MYCBP2 protein levels, miR-1247 levels, and patient survival.³³ Overall, DNA hypermethylation silences miR-1247, allowing MYCBP2 and c-myc protein to increase and promote tumor growth in CRC. So, the connection between MYCBP2, c-myc, and miR-1247 could be key in fighting tumors. Targeting this axis with demethylation

agents may offer a potential treatment option.

MiR-212

MiR-212 is an intronic miRNA located on the distal end of chromosome 17 at p13.3. It is highly conserved in vertebrates and is generated from a stable intron of a non-protein coding gene. Several studies have shown that this specific miRNA acts as a tumor suppressor and is downregulated in various types of cancers, including gastric, non-small cell, and lung cancers.³⁵ Furthermore, a separate study indicated that miR-212 could serve as a prognostic biomarker in acute myeloid leukemia.⁷⁴ The expression of miR-212 is reduced in human CRC tissues due to genetic and epigenetic factors such as promoter hypermethylation.

While MECP2, PTCH1, and PED have been previously recognized as targets of miR-212, Meng and others conducted bioinformatic analysis and experiments that revealed MnSOD as another direct target of miR-212 in CRC. They found that the 3'UTR of MnSOD serves as the functional target site for miR-212.³⁵ MnSOD is an antioxidant enzyme found in the mitochondrial matrix. It may have important implications in the development of cancer.⁷⁵ The process of EMT plays a critical role in the spread and growth of CRC cells.⁷⁶ MnSOD is a key player in this process. They used western blot analysis to measure the expression levels of MnSOD and miR-212. They discovered that a decrease in MnSOD and overexpression of miR-212 resulted in an increase in epithelial markers and a reduction of mesenchymal markers. In summary, the findings indicate that miR-212 suppresses EMT in CRC cells by repressing MnSOD activity.³⁵

The reduction of miR-212 could potentially serve as a prognostic marker for patients with CRC, as it might prevent tumor progression by targeting MnSOD messenger RNA. Both miR-212 and MnSOD could also be considered therapeutic targets for cancer treatment.

MiR-128

Takahashi and co-workers discovered that miR-128 is often reduced in advanced CRC due to increased promoter hypermethylation. Furthermore, the decrease in miR-128 levels was strongly linked to higher recurrence rates in CRC. miR-128 directly targets NEK2, causing G2-phase cell cycle arrest and suppressing cancer cell growth. Additionally, it is epigenetically silenced in CRC cells. High levels of NEK2 in CRC tissues were linked to a negative prognosis. The miR-128/NEK2 pathway could be a promising therapeutic target for individuals with CRC.⁴²

MiR-373

Tanaka and colleagues analyzed miRNA expression in CRC cell lines pre- and post-5-aza-2'-deoxycytidine (DAC) treatment. They identified 10 miRNAs with more than a 2-fold increase after DAC treatment in each cell line. Specifically, they focused on miR-373 and discovered

that its overexpression inhibited cell proliferation. Furthermore, they found that miRNA expression was repressed due to abnormal methylation in colon cancer cell lines. miR-373 serves varying roles in different types of malignant tumors, acting as a tumor suppressor in CRC. Computational predictions have identified RAB22A as a potential target gene for miR-373. In contrast, RAB22A functions as an oncogene with increased expression levels in CRC and malignant melanoma.⁷⁷ In clinical samples showing abnormal methylation of the miR-373 promoter region, the expression of miRNA was decreased, while the levels of the RAB22A target gene were elevated. This study revealed that silencing miR-373 plays a critical role in the progression of CRC.²⁶

Other miRNAs Genes

miR-133b and miR-1

MiR-133b acts as a tumor suppressor gene in CRC and is frequently silenced by CpG methylation in the promoter region.⁷⁸ Surprisingly, miR-133b actually inhibits the HOXA9/ZEB1 pathway, leading to an increase in tumor metastases and worse outcomes in CRC.⁷⁹ Furthermore, DNA hypermethylation of miR-1 was initially identified in hepatocellular carcinoma (HCC) and later found in CRC. On the other hand, miR-1 interacts with miR-133a in CRC, and silencing both microRNAs has a negative effect on TAGLN2 expression. The interaction between miR-1 and miR-133a, leading to the upregulation of TAGLN2, plays a crucial role in CRC.⁸⁰

Epigenetically Regulated Long Non-coding RNAs in CRC

LncRNAs are RNA molecules have over 200 nucleotides and cannot encode proteins^{81,82} LncRNAs play a role in various cellular processes, such as gene regulation and chromatin dynamics. They are also involved in important functions like cell proliferation, differentiation, and apoptosis.⁸³ Some reports have shown that lncRNAs play a crucial role in the development and advancement of various types of tumors.⁸⁴ We explored the epigenetically directed aberrant lncRNAs expression and their possible roles in the development and advancement of CRC (Table 2). These findings suggest that lncRNAs could serve as valuable markers for diagnosis and prognosis.

Small Nucleolar RNA Host Gene 11 (SNHG11) lncRNA

SNHG11 is an intergenic lncRNA found on the plus strand of chromosomal 20q11.23. It has a length of 4598 nt and is composed of five exons⁹⁷ SNHG11 is a key player in promoting the invasion and metastasis of CRC cells while inhibiting apoptosis. Xu and colleagues⁹³ discovered that SNHG11 lncRNAs were likely regulated by DNA methylation in The Cancer Genome Atlas (TCGA)-COAD, highlighting its significance in CRC. This suggests that DNA methylation could influence SNHG11 expression, as it is upregulated due to promoter hypomethylation in CRC. SNHG11 knockdown was

found to inhibit the migration and invasion of CRC cells under hypoxic conditions. HIF-1 α stabilization is crucial for cells adapting to changes in oxygen levels. This process is closely monitored by factors like PHD, pVHL, and ncRNAs like miR-200b, miR-200c, and miR-429. SNHG11 binds to specific sites on HIF-1 α , preventing its degradation by blocking the interaction with pVHL. Increased levels of HIF-1 α in CRC promote metastasis by controlling various target genes. Additionally, SNHG11 boosts the expression of HIF-1 α target genes like AK4, ENO1, HK2, and Twist1.^{93,98}

In summary, this finding shows that the lncRNA SNHG11 boosts the stability and activity of HIF-1 α in CRC cells, leading to increased invasion and metastasis. SNHG11 may be used as a prognostic marker and treatment target for patients with CRC.

LINC00460

Zhang and co-workers have noted that abnormal lncRNA expression significantly influences various biological processes in CRC, including tumor growth, spread, and proliferation. By analyzing the TCGA database, they pinpointed LINC00460 as the most commonly activated lncRNA in patients with CRC compared with healthy tissues.⁸⁹ LINC00460 is 9936 nucleotides long and located on the plus strand of chromosome 13q33.2.⁹⁷ It is suggested that this gene may have an oncogenic role in cancer and potentially exert a carcinogenic effect. Previous studies have demonstrated that the overexpression of LINC00460 is linked to increased cell proliferation and invasion in various types of cancer, such as gastric cancer, lung cancer, ovarian cancer, and esophageal cancer. Despite its unclear role in colon cancer, Zhang's results suggest that inhibiting the LINC00460 gene can slow down the proliferation of CRC cells, pointing to its potential carcinogenic effect on tumor growth in CRC. In conclusion, research studies in vitro and in living organisms have shown that the LINC00460 lncRNA is upregulated in CRC, triggered by DNA methylation. This gene is linked to tumor spread, promoting invasion and migration of CRC cells, which could impact patient prognosis.⁸⁹

LIFR-AS1

LIFR AS1, located on chromosome 5p13.1, is a new long ncRNA that acts as a tumor suppressor in CRC. It is transcribed in an antisense manner from the LIFR gene. Abnormal expression of LIFR-AS1 has been observed in various human tumors.⁹⁶

Zhang's research indicates that abnormal DNA methylation leads to the decreased expression of LIFR-AS1, which in turn promotes the advancement of colon cancer. They found increased methylation of a CpG island in the promoter region of LIFR-AS1, accelerating cancer progression. The methylation level in LIFR-AS1 demonstrated high sensitivity and specificity in diagnosing CRC. Functional tests conducted by Song revealed that LIFR-AS1 can competitively bind to hsa-miR-29b-

Table 2. Epigenetically regulated lncRNAs in colorectal cancer

lncRNA/ Expression pattern in CRC	Epigenetic regulation	Target gene/ signaling pathway	Oncogene or tumor- suppressor/ biomarker	Sample type	Findings	Ref
CAHM↓	DNA HypoM	-	TS/ Diagnostic	Tissue: 26 N, 21 A, 87 AC Plasma: 74 N, 73 A, 73 CRC	Methylated CAHM has been found in patients' plasma and tissue, suggesting a possible role in non-invasive CRC detection assays.	2014 ⁸⁵
TUG1↑	HDAC1	E-cadherin↑ and N-cadherin, vimentin, and Fibronectin↓	Onco/ Prognostic	120 CRC with PANS, cell lines	The high levels of TUG1 in CRC show poor prognosis, leading to lower survival rates and an increased risk of cancer metastasis. TUG1 regulates the invasive and metastatic capabilities of CRC cells, in part through the modulation of EMT.	2016 ⁸⁶
LINC00114 ↑	DNA HypoM	miR-133b	Onco/ Diagnostic	CRC cell lines	LINC00114 regulates the expression of the NUP214 protein by sponging miR-133b. LINC00114 inhibits miR-133b expression through the methylation of its promoter region by the EZH2/DNMT1 complex.	2019 ⁸⁷
H19 ↑	DNA HypoM	miR-194-5p	Onco/ Diagnostic	214 CRC with PANS, cell line	H19 inhibits miR194-5p, affecting the expression of FoxM1 and regulating the metastasis and EMT of CRC cells.	2019 ⁸⁸
LINC00460 ↑	DNA HypoM	-	Onco/ Prognostic	407 CRC tumors and 21 ANS	LINC00460 hypomethylation and expression promote CRC metastasis and are associated with poor survival rates in CRC patients.	2019 ⁸⁹
RP11 ↑	m6A methylation	SIAH1 & FBXO45	Onco/ Prognostic	CRC cell lines & tumor tissue	The expression of RP11 was significantly higher in CRC. RP11 played a crucial role in the metastasis of CRC cells by regulating Siah1-Fbxo45/Zeb1.	2019 ⁹⁰
MALAT1 ↑	DNA HypoM	β-catenin signaling pathway, AKAP-9	Onco/ Prognostic	78 CRC tissue and CRC cell lines	JMJD2C enhances the metastatic abilities of CRC cells by regulating the histone methylation level of MALAT1 promoter, thereby upregulating the expression of MALAT1 and enhancing the activity of β-catenin signaling pathway.	2019 ⁹¹
NEAT1 ↑	m6A demethylation	-	Onco/ Prognostic	70 CRC tissues and PANS	NEAT1 levels significantly increased in CRC tissues, correlated with poor prognosis. ALKBH5 facilitated the upregulation of NEAT1 expression through demethylation.	2020 ⁹²
SNHG11 ↑	DNA HypoM	HIF-1α/AK4, ENO1, HK2, and Twist1	Onco/ Prognostic	164 CRC with PANS	SNHG11 inhibits the binding of pVHL to HIF-1α by occupying the recognition sites. This action promotes migration and invasion in CRC cells by activating downstream targets of HIF-1α.	2020 ⁹³
PVT1 ↑	DNA HypoM	MYC, TGFβ/SMAD and Wnt/β-Catenin pathways	Onco/ Prognostic	426 CRC patients, CRC cell line	PVT1 enhances the oncogenic potential of MYC through epigenetic regulation. PVT1 locus could impact the expression of TGFβ/SMAD and Wnt/β-catenin pathways genes.	2020 ⁹⁴
LINC00152 ↑	DNA HypoM	Cyclin D1, PI3K/ Akt, Ras, WNT, TP53, Notch and ErbB	Onco/ Prognostic	43 N, 55 A, 43 CRC	LINC00152 significantly upregulated in CRC by promoter hypomethylation. LINC00152 contributes to CRC progression through PI3K/Akt, Ras, WNT, TP53, Notch, and ErbB.	2020 ⁹⁵
LIFR-AS1 ↓	DNA HyperM	miR-29a-3p	TS/ Diagnostic	92 CRC tissues and 43 normal tissues	DNA hypermethylation causes a decrease in LIFR-AS1, leading to the advancement of CRC. Its downregulation is associated with poor prognosis.	2022 ⁹⁶
LINC01559 ↓	DNA HypoM	miR-106b-5p	TS	Fresh CRC tissues and PANS	LINC01559 was downregulated in CRC and associated with poor prognosis. LINC01559 upregulates PTEN through sponging miR-106b-5p. LINC01559/miR-106b-5p/PTEN axis is a negative regulation of CRC.	2022 ⁸⁴

AP-2α; activator protein 2α, MDR; multidrug resistance, PANS: paired adjacent normal specimens, EMT: epithelial-to-mesenchymal transition, PANS: paired adjacent normal specimens, A: adenomas, N: normal, AC: adenocarcinoma, TS: tumor suppressor, onco: oncogene.

3p, inhibiting the proliferation, colony formation, and invasion of colon cancer cells.⁹⁶ Furthermore, Liu et al. found in a separate study that LIFR-AS1 functions as a sponge for miR-29a in CRC. Knocking down LIFR-AS1 diminished the impact of photodynamic therapy (PDT) on the proliferation and apoptosis of CRC cells, suggesting that LIFR-AS1 may act as a tumor suppressor by interacting with miR-29a.⁹⁹ Furthermore, high levels of SNRPF, which is indirectly associated with LIFR-AS1, were observed in CRC cells. Increased SNRPN expression was found to be indicative of a poor prognosis.⁹⁶ These results showed that the methylation level of LIFR-AS1 is highly sensitive and specific for diagnosing CRC and is also linked to the prognosis of the disease.

NcRNA and Drug Resistance

Although treatments like chemotherapy, targeted therapy, and immunotherapy have improved patient survival in CRC, the development of primary and secondary drug resistance poses a significant clinical challenge. The heterogeneity of CRC and the issue of drug resistance continue to hamper effective cancer treatment. Epigenetic modifications, which can be present in circulating tumor cells, play a key role in these challenges.¹⁰⁰ Therefore, targeting epigenetic regulators is now seen as a promising strategy to overcome drug resistance.¹⁰¹

The role of ncRNAs in drug resistance is increasingly recognized.¹⁰² METTL3-dependent m6A methylation of miR-181d-5p¹⁰³ by directly targeting neurocalcin δ

(NCALD) inhibits the 5-FU sensitivity of CRC cells. Also, this modification in lncRNA ADIRF-AS1 and AL139035.1 regulates 5-FU drug resistance formation through MAPK signaling.¹⁰⁴ Despite m6A methylation, DNA hypermethylation or hypomethylation in different miRNA and lncRNA have been shown to contribute to drug resistance (Table 3, Figure 2).

lncRNA Colorectal Cancer-Associated lncRNA (CCAL)

CCAL an oncogenic lncRNA, actively promotes the development and advancement of CRC. Several studies have shown that CCAL significantly contributes to the progression of various tumors, particularly CRC. Ma and others investigated the oncogenic properties and impact of CCAL on CRC. They discovered that epigenetic regulatory factors, like DNA methylation, control the expression of CCAL. Additionally, through methylation-specific PCR analysis, they observed a lower level of methylation in the CpG island region of CCAL in CRC tissue samples compared to normal tissue samples.

CCAL is one of lncRNAs that plays a crucial role in regulating molecular pathways through its interactions with proteins.^{107,113} According to Ma and colleagues, high levels of CCAL can trigger cell proliferation, invasion, cell-cycle progression, migration, and invasion in CRC by inhibiting the AP-2α protein. Additionally, CCAL activates the Wnt/β-catenin pathway by suppressing AP-2α. Conversely, reducing CCAL levels results in increased AP-2α expression, decreased β-catenin expression, and elevated levels of c-myc, cyclin D1, and E-cadherin. Furthermore, CCAL plays a role in regulating MDR1/P-gp expression by activating the Wnt signaling pathway. MDR is a significant challenge in successful chemotherapy for patients with CRC. In summary, lncRNA-CCAL controls CRC progression and MDR by activating the Wnt/β-catenin signaling pathway, suppressing AP-2α, and increasing MDR1/P-gp expression.¹⁰⁷ This finding indicates that lncRNA CCAL could serve as a valuable new prognostic biomarker for patients with CRC and advanced disease or metastasis.

Table 3. The role of epigenetically regulated non-coding RNA in drug resistance

Expression	Epigenetic	Target gene/ signaling pathway	Oncogene or Tumor-suppressor /biomarker	Sample type	Biological function	Ref
miR-148a↓	HyperM.	PXR, TGIF2, MSX1, CDC25B, DNMT1, DNMT3 and ROCK1	TS / 5-FU and oxaliplatin	273 CRC patients (76 stage II, 125 stage III, 72 stage IV)	miR-148a expression was down-regulated in advanced CRC tissues, associated with poor prognosis and poor response to 5-fluorouracil and oxaliplatin-based chemotherapy.	2012 ¹⁰⁵
miR-630 ↑	DNA HypoM	BCL2L2 and TP53RK	TS/radiosensitivity	CRC cell lines	miR-630 expression positively correlated with radiosensitivity. Methylation and CREB modulated miR-630 expression. CREB-miR-630-BCL2L2 and TP53RK pathway regulate radiosensitivity.	2015 ¹⁰⁶
CCAL↑	HypoM	AP-2α and MDR1/P-gp	Onco/ MDR	252 CRC with PANS	CCAL enhances CRC progression and multidrug resistance by activating the Wnt/β-catenin signaling through targeting AP-2α and, in turn, MDR1/P-gp, respectively. Patients with high CCAL expression show shorter overall survival and worse response to adjuvant chemotherapy.	2016 ¹⁰⁷
MIR100HG↑ miR-100 ↑ miR-125b↑	HypoM	Wnt/β-catenin negative regulators	Onco / cetuximab-resistant	CRC cell lines	MIR100HG-derived miR-100 and miR-125b mediate cetuximab resistance via Wnt/β-catenin pathway. GATA6 represses MIR100HG, but this repression is relieved by miR-125b targeting of GATA6.	2017 ¹⁰⁸
miR-181a↓ miR-135a↓ miR-302c↓	HyperM	PLAG1/IGF2 signaling	TS / 5-FU	67 CRC, and cell lines	miR-181a/135a/302c function as tumor suppressors via repressing PLAG1/IGF2 signaling. Their expression promoted the sensitivity of CRC cells to 5-FU treatment.	2018 ¹⁰⁹
MEG3↓	HyperM ¹¹⁰	miR-141/ PDCD4	TS/oxaliplatin	48 CRC with PANS, cell lines	Low MEG3 expression was correlated with poor prognosis. MEG3 was down-regulated in oxaliplatin-resistant CRC tissues and cell lines. MEG3 elevated PDCD4 expression through targeting miR-141	2018 ¹¹¹
miR-34a↓	HyperM	CSF1R, SNAIL	TS / 5-FU	CRC cell lines	CpG-methylation of miR-34a results in elevated expression of CSF1R and 5-FU resistance. High CSF1R expression is associated with poor prognosis and metastasis.	2020 ⁶³
miR-149 ↓	HyperM	Akt, cyclin B1, CDK	TS / MDR	CRC cell lines	Hypomethylation of the miR-149 CpG island and upregulation triggers cell cycle arrest by reducing the expression of AKT, Cyclin B1, and CDK1. Thus, leads to improved sensitivity to chemotherapy in CRC.	2021 ¹¹²
miR-181d-5p	m6A methylation	NCALD	TS / 5-FU	141 CRC tissues and FFPE	METTL3-dependent m6A methylation was upregulated in CRC to promote the processing of miR-181d-5p. This led to increased miR-181d-5p expression, which inhibited the 5-FU sensitivity of CRC cells by targeting NCALD.	2022 ¹⁰³
ADIRF-AS1 AL139035.1	m6A methylation	MAPK signaling	onco / 5-FU	CRC cell lines	ADIRF-AS1 and AL139035.1 promote CRC progression and may regulate drug resistance through MAPK signaling (FOS, DUSP1, MEK2C).	2024 ¹⁰⁴

MDR: Multidrug resistance; EMT: epithelial-to-mesenchymal transition; PANS: paired adjacent normal specimens; A: adenomas; N: normal.

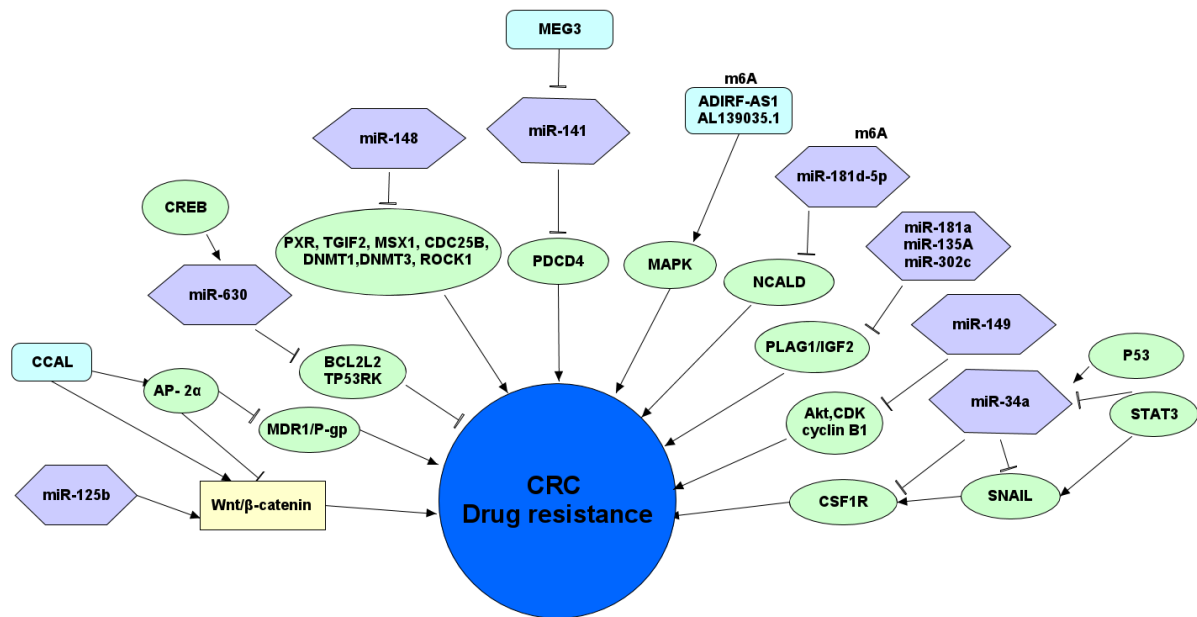


Figure 2. Role of epigenetically regulated non-coding RNAs in drug resistance in colorectal cancer

MiRNA-125

The miR-125 family consists of two distinct members, miR-125a and miR-125b, which are located on separate chromosomes. The miR-125a molecule consists of 86 bases and is found on chromosome 19q13.41 on the plus strand. In contrast, miR-125b is an 88-base molecule located on chromosome 11q24.1 on the minus strand.⁴⁶ Recent studies show that the miR-125 family is dysregulated in various types of human cancer, including gliomas, prostate cancer, breast cancer, and gastric cancer. Depending on the type of cell, both miR-125a and miR-125b can either promote cancer growth or suppress it. For example, in prostate cancer, miR-125b acts as an oncogene and promotes tumor growth by inhibiting the intrinsic apoptosis pathway by targeting PUMA, P53, and BAK.¹¹⁴ In contrast, miR-125b acts as a tumor suppressor in breast cancer by suppressing the oncoproteins MUC1, ERBB2, and ERBB3, thereby inhibiting tumor growth.⁴⁵ However, miR-125a significantly reduced the growth, movement, and infiltration of cancer cells, including gastric and breast cancer^{45,115} While the role of miR-125 in CRC is not yet fully understood, a recent study revealed that both miR-125a and b are frequently reduced in CRC tissues by hypermethylation. This suggests that the miR-125 family may possess tumor-suppressing properties in CRC⁴⁶ (Table 1). However, hypomethylation of lncRNA MIR100HG region and, consequently upregulation of miR-125b associated with cetuximab resistance via Wnt/ β -catenin pathway¹⁰⁸ (Table 3), and depression of miR-125b-2-3p associated with cell sensitivity to first-line chemotherapy (fluorouracil, oxaliplatin, CPT-11).¹¹⁶

MiR-34

The miR-34 family consists of three members, namely miR-34a, miR-34b, and miR-34c, which are encoded by

genes located on chromosomes 1 and 11. These miR-34 family members exhibit tumor suppressor properties by suppressing the expression of their target mRNAs.¹¹⁷ The regulation of miR-34 expression involves various mechanisms that contribute to its dysregulation in cancer. Studies have shown reduced expression of miR-34b/c,²⁴ miR-34c-5p,⁵⁵ and miR-34a^{30,63} in CRC through epigenetic mechanisms. The tumor suppressor protein p53 directly interacts with the miR-34 gene promoter, leading to the activation of its transcription. Specifically, the activation of miR-34a enhances the functions of p53, including cell cycle arrest, DNA repair, and apoptosis.¹¹⁸ Furthermore, TP53 gene polymorphisms have been linked to the methylation and expression levels of miR-34a/b/c in CRC tissues.¹¹⁹

The miR-34 gene is targeted by activated STAT3, repressing miR-34 transcription and promoting EMT in CRC cells and tumors.¹¹⁷ Additionally, in CRC, the control of tyrosine kinase colony-stimulating factor 1 receptor (CSF1R) by p53-inducible miR-34a is disrupted due to a feedback loop involving STAT3. Shi and colleagues⁶³ showed that miR-34a directly impacts CSF1R and plays a crucial role in the collaborative action of p53 and miR-34a in limiting CRC progression. P53 reduces CSF1R expression by upregulating miR-34a, while SNAIL increases CSF1R expression by downregulating miR-34a both directly and indirectly. CSF1R, when activated through a STAT3-mediated pathway, promotes EMT, migration, colonization, and metastasis in CRC cells. Methylation of CpG sites on miR-34a leads to increased expression of CSF1R, contributing to resistance to 5-FU in CRC cells (Table 3).

lncRNAs Act as Competing Endogenous RNA (ceRNAs)

In 2011, Pier Paolo Pandolfi's group introduced the

concept of a new RNA interaction mechanism known as ceRNA. This theory proposes that various types of RNAs, including coding RNAs and ncRNAs (like lncRNAs, circRNAs, and pseudogenes), communicate with each other through miRNA complementary sequences called MREs. This interplay creates a vast regulatory network within the transcriptome. Many lncRNAs play a key role in regulating gene expression by interacting with microRNAs through a process known as ceRNA mechanism.¹²⁰ Previous research demonstrated that LIFR-AS1 functions as a ceRNA in various types of cancer. Specifically, LIFR-AS1 has been found to sponge miR-29a-3p and miR-4698 in gastric cancer, miR-150-5p in pancreatic cancer, miR-942-5p in lung cancer, miR-197-3p in breast cancer, miR-4262 in glioma and miR-31-5p in thyroid carcinoma.⁹⁶

In a study by Lin et al, LIFR-AS1 was identified as a ceRNA for miR-29a, which inhibits its expression and increases TNFAIP3 expression. This process helps regulate resistance to PDT in CRC. The researchers observed a negative regulatory relationship between LIFR-AS1 and miR-29a in PDT-treated HCT116 cells through direct binding. Knocking down LIFR-AS1 reduced the impact of PDT on CRC cell proliferation and apoptosis, suggesting LIFR-AS1 may function as a tumor suppressor by interacting with miR-29a.⁵⁴ In addition, Song and colleagues conducted a study on the ceRNA function of LIFR-AS1 in CRC. They discovered that LIFR-AS1 can interact with hsa-miR-29b-3p using a luciferase reporter gene in colon cancer cells.⁹⁶

H19 is an oncofetal ncRNA that is hypomethylated and upregulated in CRC, promoting its development by generating miRNA or serving as ceRNA.¹²¹ H19 and miR-194-5p alter the EMT, invasion, and migration of CRC cells by targeting downstream FoxM1. FoxM1, influenced by H19 and miR-194, serves as an oncogene in CRC. H19 can regulate EMT-related genes by sponging miRNAs. In addition, FoxM1 can counteract the effects of miR-194-5p on suppressing invasion, migration, and EMT in CRC cells. Li and co-workers demonstrated the lncRNA H19/miR-194/FoxM1 axes could be a valuable target for diagnosing and treating CRC.⁸⁸

The lncRNA LINC00114 is associated with cancer lncRNA and is upregulated in CRC. Through DNA methylation, LINC00114 negatively regulates the expression of miR-133b, indicating its role as an oncogene in CRC development. Research has shown that miR-133b is crucial in advancing CRC as it inhibits cell growth and spread. NUP214 plays a crucial role in mitosis and cancer development, and it has been identified as a direct target of miR-133b. A study by Lv and others showed that LINC00114 can regulate the expression of the NUP214 protein by acting as a sponge for miR-133b.⁸⁷

snoRNAs

snoRNAs are a crucial class of ncRNAs that may undergo changes in human cancer. These RNAs are located in the

nucleolus and play a significant role in various cellular functions, including RNA modification, pre-RNA processing, and the regulation of alternative splicing. Studies have suggested that snoRNAs could contribute to the development and progression of cancer. Ferreira and co-workers conducted research using Bisulfite genomic sequencing on multiple clones from normal colon mucosa and the CRC cell line hCT-116, revealing that certain snoRNAs were hypomethylated while others were hypermethylated.

In cancer cells, snoRNAs SNORD123, U70c, and Aca59B, as well as the 5'-cpG islands associated with their host genes, were hypermethylated, which was not observed in the corresponding normal tissue. Recent research has shown that snoRNAs are frequently hypermethylated in different tumors, specifically in leukemias and CRC. This highlights the need for a more in-depth investigation of this specific group of ncRNAs that are affected by epigenetic changes in human cancer.¹²²

RNA Epitranscriptome

Various chemical modifications occur on RNA bases and ribose molecules, playing a crucial role in the post-transcriptional regulation of gene expression. To date, various types of RNA modifications have been identified on both coding and predominantly ncRNA molecules. Similar to modifications found on DNA and histone proteins, RNA modifications can be added, removed, and recognized by specific enzymes. These modifications typically impact RNA processes such as splicing, stability, localization, translation, and interactions between RNA molecules and RNA-binding proteins, thereby influencing cellular activities.¹²³

Recent studies have highlighted the emerging role of RNA modifications in various cancers, including CRC.^{124,125} Substantial evidence demonstrates the impact of m6A modification on the progression and development of drug resistance in CRC (Tables 2 and 3). One example of this is the upregulated expression of RP11 in CRC, which has been associated with m6A modification, leading to its localization to chromatin. The upregulation of RP11 stimulates the expression of Zeb1 by downregulating Siah1 and Fbxo45 mRNA expression as RP11 binds to hnRNPA2B1. This mechanism ultimately results in the degradation of Siah1 and Fbxo45, thereby preventing the degradation of the mesenchymal transition-related gene Zeb1. Zeb1, functioning as an epithelial-mesenchymal transition transcription factor (EMT-TF), plays a critical role in promoting EMT progression by specifically targeting E-Cadherin expression.⁹⁰

Additionally, elevated levels of NEAT1 have been observed in CRC tissues and are associated with a poor prognosis. The upregulation of NEAT1 expression is mediated by ALKBH5 through m6A demethylation.⁹² NEAT1 plays a role in CRC advancement by sponging miR-193a-3p and interacting with DDX5, thereby influencing KRAS expression and Wnt/ β -catenin

signaling.¹²⁶

Moreover, CRC shows an upregulation of METTL3-dependent m6A methylation, which promotes the processing of miR-181d-5p. This results in increased expression of miR-181d-5p, leading to reduced sensitivity of CRC cells to 5-FU by targeting NCALD.¹⁰³ The m6A methylation of lncRNAs has been identified in 5FU-resistant HCT15 cells, suggesting a role in regulating mRNA expression of drug resistance-associated genes and promoting cancer progression. In particular, the silencing of two specific lncRNAs, ADIRF-AS1 and AL139035.1, associated with MAPK signaling pathways involving FOS, DUSP1, and MEF2C genes has been found to enhance proliferation, metastasis, and potentially regulating drug resistance through.¹⁰⁴

Conclusion

Extensive research has been dedicated to uncovering the molecular pathology of CRC and developing novel epigenetic biomarker assays for accurately diagnosing and predicting the prognosis of this disease. The substantial impact of epigenetic modifications on the onset and advancement of CRC has driven this research focus. Recent findings suggest that abnormal epigenetic alterations and the dysregulation of ncRNAs, including miRNAs and lncRNAs, offer promising avenues for serving as biomarkers in CRC. These biomarkers have the potential to contribute to early detection, prognosis determination, and the identification of therapeutic targets. Further exploration is necessary to fully grasp the role of ncRNA epigenetics in the development of CRC and assess its viability as a diagnostic or prognostic tool for managing CRC effectively.

Authors' Contribution

Conceptualization: Majid Zaki-dizaji.

Data curation: Zahra Taheri.

Supervision: Majid Zaki-dizaji.

Visualization: Zahra Taheri.

Writing—original draft: Zahra Taheri.

Writing—review & editing: Majid Zaki-dizaji.

Competing Interests

The authors declare no conflict of interest related to this work.

Ethical Approval

Not applicable.

Funding

No funding was received for this study.

References

- Fazeli MS, Keramati MR. Rectal cancer: a review. *Med J Islam Repub Iran* 2015;29:171.
- Li Z, Tan H, Yu H, Deng Z, Zhou X, Wang M. DNA methylation and gene expression profiles characterize epigenetic regulation of lncRNAs in colon adenocarcinoma. *J Cell Biochem* 2020;121(3):2406-15. doi: [10.1002/jcb.29463](https://doi.org/10.1002/jcb.29463)
- Wang R, Su Q, Yan ZP. Reconsideration of recurrence and metastasis in colorectal cancer. *World J Clin Cases* 2021;9(24):6964-8. doi: [10.12998/wjcc.v9.i24.6964](https://doi.org/10.12998/wjcc.v9.i24.6964)
- Razzaghi H, Khabbazzpour M, Heidary Z, Heiat M, Shirzad Moghaddam Z, Derogar P, et al. Emerging role of tumor-educated platelets as a new liquid biopsy tool for colorectal cancer. *Arch Iran Med* 2023;26(8):447-54. doi: [10.34172/aim.2023.68](https://doi.org/10.34172/aim.2023.68)
- Khabbazzpour M, Tat M, Karbasi A, Abyazi MA, Khodadoustan G, Heidary Z, et al. Advances in blood DNA methylation-based assay for colorectal cancer early detection: a systematic updated review. *Gastroenterol Hepatol Bed Bench* 2024;17(3):225-40. doi: [10.22037/ghfbb.v17i3.2978](https://doi.org/10.22037/ghfbb.v17i3.2978)
- Oh CK, Cho YS. Pathogenesis and biomarkers of colorectal cancer by epigenetic alteration. *Intest Res* 2024;22(2):131-51. doi: [10.5217/ir.2023.00115](https://doi.org/10.5217/ir.2023.00115)
- Yu X, Zhao H, Wang R, Chen Y, Ouyang X, Li W, et al. Cancer epigenetics: from laboratory studies and clinical trials to precision medicine. *Cell Death Discov* 2024;10(1):28. doi: [10.1038/s41420-024-01803-z](https://doi.org/10.1038/s41420-024-01803-z)
- Yang J, Xu J, Wang W, Zhang B, Yu X, Shi S. Epigenetic regulation in the tumor microenvironment: molecular mechanisms and therapeutic targets. *Signal Transduct Target Ther* 2023;8(1):210. doi: [10.1038/s41392-023-01480-x](https://doi.org/10.1038/s41392-023-01480-x)
- Xu M, Chen X, Lin K, Zeng K, Liu X, Pan B, et al. The long noncoding RNA SNHG1 regulates colorectal cancer cell growth through interactions with EZH2 and miR-154-5p. *Mol Cancer* 2018;17(1):141. doi: [10.1186/s12943-018-0894-x](https://doi.org/10.1186/s12943-018-0894-x)
- Vos PD, Leedman PJ, Filipovska A, Rackham O. Modulation of miRNA function by natural and synthetic RNA-binding proteins in cancer. *Cell Mol Life Sci* 2019;76(19):3745-52. doi: [10.1007/s00018-019-03163-9](https://doi.org/10.1007/s00018-019-03163-9)
- Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs and its biological functions. *Nat Rev Mol Cell Biol* 2021;22(2):96-118. doi: [10.1038/s41580-020-00315-9](https://doi.org/10.1038/s41580-020-00315-9)
- Cech TR, Steitz JA. The noncoding RNA revolution-trashing old rules to forge new ones. *Cell* 2014;157(1):77-94. doi: [10.1016/j.cell.2014.03.008](https://doi.org/10.1016/j.cell.2014.03.008)
- Beermann J, Piccoli MT, Viereck J, Thum T. Non-coding RNAs in development and disease: background, mechanisms, and therapeutic approaches. *Physiol Rev* 2016;96(4):1297-325. doi: [10.1152/physrev.00041.2015](https://doi.org/10.1152/physrev.00041.2015)
- Hombach S, Kretz M. Non-coding RNAs: classification, biology and functioning. *Adv Exp Med Biol* 2016;937:3-17. doi: [10.1007/978-3-319-42059-2_1](https://doi.org/10.1007/978-3-319-42059-2_1)
- Jia Z, An J, Liu Z, Zhang F. Non-coding RNAs in colorectal cancer: their functions and mechanisms. *Front Oncol* 2022;12:783079. doi: [10.3389/fonc.2022.783079](https://doi.org/10.3389/fonc.2022.783079)
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116(2):281-97. doi: [10.1016/s0092-8674\(04\)00045-5](https://doi.org/10.1016/s0092-8674(04)00045-5)
- Fu G, Brkić J, Hayder H, Peng C. MicroRNAs in human placental development and pregnancy complications. *Int J Mol Sci* 2013;14(3):5519-44. doi: [10.3390/ijms14035519](https://doi.org/10.3390/ijms14035519)
- Muhammad S, Kaur K, Huang R, Zhang Q, Kaur P, Yazdani HO, et al. MicroRNAs in colorectal cancer: role in metastasis and clinical perspectives. *World J Gastroenterol* 2014;20(45):17011-9. doi: [10.3748/wjg.v20.i45.17011](https://doi.org/10.3748/wjg.v20.i45.17011)
- Svoronos AA, Engelman DM, Slack FJ. OncomiR or tumor suppressor? The duplicity of microRNAs in cancer. *Cancer Res* 2016;76(13):3666-70. doi: [10.1158/0008-5472.Can-16-0359](https://doi.org/10.1158/0008-5472.Can-16-0359)
- Tüfekci KU, Oner MG, Meuwissen RL, Genç S. The role of microRNAs in human diseases. *Methods Mol Biol* 2014;1107:33-50. doi: [10.1007/978-1-62703-748-8_3](https://doi.org/10.1007/978-1-62703-748-8_3)
- Paul P, Chakraborty A, Sarkar D, Langthasa M, Rahman M, Bari M, et al. Interplay between miRNAs and human diseases. *J Cell Physiol* 2018;233(3):2007-18. doi: [10.1002/jcp.25854](https://doi.org/10.1002/jcp.25854)
- Lujambio A, Ropero S, Ballestar E, Fraga MF, Cerrato C, Setièn F, et al. Genetic unmasking of an epigenetically

- silenced microRNA in human cancer cells. *Cancer Res* 2007;67(4):1424-9. doi: [10.1158/0008-5472.Can-06-4218](https://doi.org/10.1158/0008-5472.Can-06-4218)
23. Grady WM, Parkin RK, Mitchell PS, Lee JH, Kim YH, Tsuchiya KD, et al. Epigenetic silencing of the intronic microRNA hsa-miR-342 and its host gene EVL in colorectal cancer. *Oncogene* 2008;27(27):3880-8. doi: [10.1038/onc.2008.10](https://doi.org/10.1038/onc.2008.10)
 24. Toyota M, Suzuki H, Sasaki Y, Maruyama R, Imai K, Shinomura Y, et al. Epigenetic silencing of microRNA-34b/c and B-cell translocation gene 4 is associated with CpG island methylation in colorectal cancer. *Cancer Res* 2008;68(11):4123-32. doi: [10.1158/0008-5472.Can-08-0325](https://doi.org/10.1158/0008-5472.Can-08-0325)
 25. Tang JT, Wang JL, Du W, Hong J, Zhao SL, Wang YC, et al. MicroRNA 345, a methylation-sensitive microRNA is involved in cell proliferation and invasion in human colorectal cancer. *Carcinogenesis* 2011;32(8):1207-15. doi: [10.1093/carcin/bgr114](https://doi.org/10.1093/carcin/bgr114)
 26. Tanaka T, Arai M, Wu S, Kanda T, Miyauchi H, Imazeki F, et al. Epigenetic silencing of microRNA-373 plays an important role in regulating cell proliferation in colon cancer. *Oncol Rep* 2011;26(5):1329-35. doi: [10.3892/or.2011.1401](https://doi.org/10.3892/or.2011.1401)
 27. Wang F, Ma YL, Zhang P, Shen TY, Shi CZ, Yang YZ, et al. SP1 mediates the link between methylation of the tumour suppressor miR-149 and outcome in colorectal cancer. *J Pathol* 2013;229(1):12-24. doi: [10.1002/path.4078](https://doi.org/10.1002/path.4078)
 28. Liu C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, Li H, et al. The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Nat Med* 2011;17(2):211-5. doi: [10.1038/nm.2284](https://doi.org/10.1038/nm.2284)
 29. Kaller M, Liffers ST, Oeljeklaus S, Kuhlmann K, Röh S, Hoffmann R, et al. Genome-wide characterization of miR-34a induced changes in protein and mRNA expression by a combined pulsed SILAC and microarray analysis. *Mol Cell Proteomics* 2011;10(8):M111.010462. doi: [10.1074/mcp.M111.010462](https://doi.org/10.1074/mcp.M111.010462)
 30. Siemens H, Neumann J, Jackstadt R, Mansmann U, Horst D, Kirchner T, et al. Detection of miR-34a promoter methylation in combination with elevated expression of c-Met and β -catenin predicts distant metastasis of colon cancer. *Clin Cancer Res* 2013;19(3):710-20. doi: [10.1158/1078-0432.Ccr-12-1703](https://doi.org/10.1158/1078-0432.Ccr-12-1703)
 31. Guo ST, Jiang CC, Wang GP, Li YP, Wang CY, Guo XY, et al. MicroRNA-497 targets insulin-like growth factor 1 receptor and has a tumour suppressive role in human colorectal cancer. *Oncogene* 2013;32(15):1910-20. doi: [10.1038/onc.2012.214](https://doi.org/10.1038/onc.2012.214)
 32. Liu L, Chen L, Xu Y, Li R, Du X. MicroRNA-195 promotes apoptosis and suppresses tumorigenicity of human colorectal cancer cells. *Biochem Biophys Res Commun* 2010;400(2):236-40. doi: [10.1016/j.bbrc.2010.08.046](https://doi.org/10.1016/j.bbrc.2010.08.046)
 33. Menigatti M, Staiano T, Manser CN, Bauerfeind P, Komljenovic A, Robinson M, et al. Epigenetic silencing of monoallelically methylated miRNA loci in precancerous colorectal lesions. *Oncogenesis* 2013;2(7):e56. doi: [10.1038/onscis.2013.21](https://doi.org/10.1038/onscis.2013.21)
 34. Ye J, Wu X, Wu D, Wu P, Ni C, Zhang Z, et al. miRNA-27b targets vascular endothelial growth factor C to inhibit tumor progression and angiogenesis in colorectal cancer. *PLoS One* 2013;8(4):e60687. doi: [10.1371/journal.pone.0060687](https://doi.org/10.1371/journal.pone.0060687)
 35. Meng X, Wu J, Pan C, Wang H, Ying X, Zhou Y, et al. Genetic and epigenetic down-regulation of microRNA-212 promotes colorectal tumor metastasis via dysregulation of MnSOD. *Gastroenterology* 2013;145(2):426-36.e6. doi: [10.1053/j.gastro.2013.04.004](https://doi.org/10.1053/j.gastro.2013.04.004)
 36. Zhang Y, Wang X, Xu B, Wang B, Wang Z, Liang Y, et al. Epigenetic silencing of miR-126 contributes to tumor invasion and angiogenesis in colorectal cancer. *Oncol Rep* 2013;30(4):1976-84. doi: [10.3892/or.2013.2633](https://doi.org/10.3892/or.2013.2633)
 37. Xia Y, Wu Y, Liu B, Wang P, Chen Y. Downregulation of miR-638 promotes invasion and proliferation by regulating SOX2 and induces EMT in NSCLC. *FEBS Lett* 2014;588(14):2238-45. doi: [10.1016/j.febslet.2014.05.002](https://doi.org/10.1016/j.febslet.2014.05.002)
 38. Ma K, Pan X, Fan P, He Y, Gu J, Wang W, et al. Loss of miR-638 in vitro promotes cell invasion and a mesenchymal-like transition by influencing SOX2 expression in colorectal carcinoma cells. *Mol Cancer* 2014;13:118. doi: [10.1186/1476-4598-13-118](https://doi.org/10.1186/1476-4598-13-118)
 39. Zhao LY, Yao Y, Han J, Yang J, Wang XF, Tong DD, et al. miR-638 suppresses cell proliferation in gastric cancer by targeting Sp2. *Dig Dis Sci* 2014;59(8):1743-53. doi: [10.1007/s10620-014-3087-5](https://doi.org/10.1007/s10620-014-3087-5)
 40. Zhang J, Fei B, Wang Q, Song M, Yin Y, Zhang B, et al. MicroRNA-638 inhibits cell proliferation, invasion and regulates cell cycle by targeting tetraspanin 1 in human colorectal carcinoma. *Oncotarget* 2014;5(23):12083-96. doi: [10.18632/oncotarget.2499](https://doi.org/10.18632/oncotarget.2499)
 41. Yin Y, Zhang B, Wang W, Fei B, Quan C, Zhang J, et al. miR-204-5p inhibits proliferation and invasion and enhances chemotherapeutic sensitivity of colorectal cancer cells by downregulating RAB22A. *Clin Cancer Res* 2014;20(23):6187-99. doi: [10.1158/1078-0432.Ccr-14-1030](https://doi.org/10.1158/1078-0432.Ccr-14-1030)
 42. Takahashi Y, Iwaya T, Sawada G, Kurashige J, Matsumura T, Uchi R, et al. Up-regulation of NEK2 by microRNA-128 methylation is associated with poor prognosis in colorectal cancer. *Ann Surg Oncol* 2014;21(1):205-12. doi: [10.1245/s10434-013-3264-3](https://doi.org/10.1245/s10434-013-3264-3)
 43. Zheng YB, Luo HP, Shi Q, Hao ZN, Ding Y, Wang QS, et al. miR-132 inhibits colorectal cancer invasion and metastasis via directly targeting ZEB2. *World J Gastroenterol* 2014;20(21):6515-22. doi: [10.3748/wjg.v20.i21.6515](https://doi.org/10.3748/wjg.v20.i21.6515)
 44. Qin J, Ke J, Xu J, Wang F, Zhou Y, Jiang Y, et al. Downregulation of microRNA-132 by DNA hypermethylation is associated with cell invasion in colorectal cancer. *Oncotargets Ther* 2015;8:3639-48. doi: [10.2147/ott.S91560](https://doi.org/10.2147/ott.S91560)
 45. Scott GK, Goga A, Bhaumik D, Berger CE, Sullivan CS, Benz CC. Coordinate suppression of ERBB2 and ERBB3 by enforced expression of micro-RNA miR-125a or miR-125b. *J Biol Chem* 2007;282(2):1479-86. doi: [10.1074/jbc.M609383200](https://doi.org/10.1074/jbc.M609383200)
 46. Chen H, Xu Z. Hypermethylation-associated silencing of miR-125a and miR-125b: a potential marker in colorectal cancer. *Dis Markers* 2015;2015:345080. doi: [10.1155/2015/345080](https://doi.org/10.1155/2015/345080)
 47. Lv LV, Zhou J, Lin C, Hu G, Yi LU, Du J, et al. DNA methylation is involved in the aberrant expression of miR-133b in colorectal cancer cells. *Oncol Lett* 2015;10(2):907-12. doi: [10.3892/ol.2015.3336](https://doi.org/10.3892/ol.2015.3336)
 48. Yu FY, Tu Y, Deng Y, Guo C, Ning J, Zhu Y, et al. MiR-4500 is epigenetically downregulated in colorectal cancer and functions as a novel tumor suppressor by regulating HMGA2. *Cancer Biol Ther* 2016;17(11):1149-57. doi: [10.1080/15384047.2016.1235661](https://doi.org/10.1080/15384047.2016.1235661)
 49. Salvi A, Sabelli C, Moncini S, Venturin M, Arici B, Riva P, et al. MicroRNA-23b mediates urokinase and c-Met downmodulation and a decreased migration of human hepatocellular carcinoma cells. *FEBS J* 2009;276(11):2966-82. doi: [10.1111/j.1742-4658.2009.07014.x](https://doi.org/10.1111/j.1742-4658.2009.07014.x)
 50. Rogler CE, Levoci L, Ader T, Massimi A, Tchaikovskaya T, Norel R, et al. MicroRNA-23b cluster microRNAs regulate transforming growth factor-beta/bone morphogenetic protein signaling and liver stem cell differentiation by targeting Smads. *Hepatology* 2009;50(2):575-84. doi: [10.1002/hep.22982](https://doi.org/10.1002/hep.22982)
 51. Kou CH, Zhou T, Han XL, Zhuang HJ, Qian HX. Downregulation of mir-23b in plasma is associated with poor prognosis in patients with colorectal cancer. *Oncol Lett* 2016;12(6):4838-44. doi: [10.3892/ol.2016.5265](https://doi.org/10.3892/ol.2016.5265)
 52. Liang J, Zhou W, Sakre N, DeVecchio J, Ferrandon S, Ting AH, et al. Epigenetically regulated miR-1247 functions as a novel tumour suppressor via MYCBP2 in methylator colon cancers. *Br J Cancer* 2018;119(10):1267-77. doi: [10.1038/s41416-018-0249-9](https://doi.org/10.1038/s41416-018-0249-9)

53. Selcuklu SD, Donoghue MT, Spillane C. miR-21 as a key regulator of oncogenic processes. *Biochem Soc Trans* 2009;37(Pt 4):918-25. doi: [10.1042/bst0370918](#)
54. Cheng YW, Chou CJ, Yang PM. Ten-eleven translocation 1 (TET1) gene is a potential target of miR-21-5p in human colorectal cancer. *Surg Oncol* 2018;27(1):76-81. doi: [10.1016/j.suronc.2017.12.004](#)
55. Gu J, Wang G, Liu H, Xiong C. SATB2 targeted by methylated miR-34c-5p suppresses proliferation and metastasis attenuating the epithelial-mesenchymal transition in colorectal cancer. *Cell Prolif* 2018;51(4):e12455. doi: [10.1111/cpr.12455](#)
56. Kashani E, Hadizadeh M, Chaleshi V, Mirfakhraie R, Young C, Savabkar S, et al. The differential DNA hypermethylation patterns of microRNA-137 and microRNA-342 locus in early colorectal lesions and tumours. *Biomolecules* 2019;9(10):519. doi: [10.3390/biom9100519](#)
57. Park SM, Gaur AB, Lengyel E, Peter ME. The miR-200 family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors ZEB1 and ZEB2. *Genes Dev* 2008;22(7):894-907. doi: [10.1101/gad.1640608](#)
58. Taheri Z, Asadzadeh Aghdaei H, Irani S, Modarressi MH, Noormohammadi Z. Clinical correlation of miR-200c/141 cluster DNA methylation and miR-141 expression with the clinicopathological features of colorectal primary lesions/tumors. *Rep Biochem Mol Biol* 2019;8(3):208-15.
59. Fessler E, Jansen M, De Sousa EMF, Zhao L, Prasetyanti PR, Rodermond H, et al. A multidimensional network approach reveals microRNAs as determinants of the mesenchymal colorectal cancer subtype. *Oncogene* 2016;35(46):6026-37. doi: [10.1038/ncr.2016.134](#)
60. Wang C, Ma X, Zhang J, Jia X, Huang M. DNMT1 maintains the methylation of miR-152-3p to regulate TMSB10 expression, thereby affecting the biological characteristics of colorectal cancer cells. *IUBMB Life* 2020;72(11):2432-43. doi: [10.1002/iub.2366](#)
61. Silber J, Jacobsen A, Ozawa T, Harinath G, Pedraza A, Sander C, et al. miR-34a repression in proneural malignant gliomas upregulates expression of its target PDGFRA and promotes tumorigenesis. *PLoS One* 2012;7(3):e33844. doi: [10.1371/journal.pone.0033844](#)
62. Garofalo M, Jeon YJ, Nuovo GJ, Middleton J, Secchiero P, Joshi P, et al. MiR-34a/c-dependent PDGFR- α/β downregulation inhibits tumorigenesis and enhances TRAIL-induced apoptosis in lung cancer. *PLoS One* 2013;8(6):e67581. doi: [10.1371/journal.pone.0067581](#)
63. Shi X, Kaller M, Rokavec M, Kirchner T, Horst D, Hermeking H. Characterization of a p53/miR-34a/CSF1R/STAT3 feedback loop in colorectal cancer. *Cell Mol Gastroenterol Hepatol* 2020;10(2):391-418. doi: [10.1016/j.jcmgh.2020.04.002](#)
64. Lee MR, Kim JS, Kim KS. miR-124a is important for migratory cell fate transition during gastrulation of human embryonic stem cells. *Stem Cells* 2010;28(9):1550-9. doi: [10.1002/stem.490](#)
65. Cheng LC, Pastrana E, Tavazoie M, Doetsch F. miR-124 regulates adult neurogenesis in the subventricular zone stem cell niche. *Nat Neurosci* 2009;12(4):399-408. doi: [10.1038/nn.2294](#)
66. Zhou L, Xu Z, Ren X, Chen K, Xin S. MicroRNA-124 (miR-124) inhibits cell proliferation, metastasis and invasion in colorectal cancer by downregulating rho-associated protein kinase 1 (ROCK1). *Cell Physiol Biochem* 2016;38(5):1785-95. doi: [10.1159/000443117](#)
67. Lujambio A. CpG island hypermethylation of tumor suppressor microRNAs in human cancer. *Cell Cycle* 2007;6(12):1454-8. doi: [10.4161/cc.6.12.4408](#)
68. Harada T, Yamamoto E, Yamano HO, Nojima M, Maruyama R, Kumegawa K, et al. Analysis of DNA methylation in bowel lavage fluid for detection of colorectal cancer. *Cancer Prev Res (Phila)* 2014;7(10):1002-10. doi: [10.1158/1940-6207.Capr-14-0162](#)
69. Otsubo T, Akiyama Y, Hashimoto Y, Shimada S, Goto K, Yuasa Y. MicroRNA-126 inhibits SOX2 expression and contributes to gastric carcinogenesis. *PLoS One* 2011;6(1):e16617. doi: [10.1371/journal.pone.0016617](#)
70. Stoeltzing O, Liu W, Reinmuth N, Parikh A, Ahmad SA, Jung YD, et al. New approaches to the treatment of hepatic malignancies angiogenesis and antiangiogenic therapy of colon cancer liver metastasis. *Ann Surg Oncol* 2003;10(7):722-33. doi: [10.1245/aso.2003.07.019](#)
71. Li XM, Wang AM, Zhang J, Yi H. Down-regulation of miR-126 expression in colorectal cancer and its clinical significance. *Med Oncol* 2011;28(4):1054-7. doi: [10.1007/s12032-010-9637-6](#)
72. Guo L, Yang S, Zhao Y, Zhang H, Wu Q, Chen F. Global analysis of miRNA gene clusters and gene families reveals dynamic and coordinated expression. *Biomed Res Int* 2014;2014:782490. doi: [10.1155/2014/782490](#)
73. Holland S, Scholich K. Regulation of neuronal functions by the E3-ubiquitinligase protein associated with MYC (MYCBP2). *Commun Integr Biol* 2011;4(5):513-5. doi: [10.4161/cib.4.5.15967](#)
74. Sun SM, Rockova V, Bullinger L, Dijkstra MK, Döhner H, Löwenberg B, et al. The prognostic relevance of miR-212 expression with survival in cytogenetically and molecularly heterogeneous AML. *Leukemia* 2013;27(1):100-6. doi: [10.1038/leu.2012.158](#)
75. Hu Y, Rosen DG, Zhou Y, Feng L, Yang G, Liu J, et al. Mitochondrial manganese-superoxide dismutase expression in ovarian cancer: role in cell proliferation and response to oxidative stress. *J Biol Chem* 2005;280(47):39485-92. doi: [10.1074/jbc.M503296200](#)
76. Thiery JP. Epithelial-mesenchymal transitions in development and pathologies. *Curr Opin Cell Biol* 2003;15(6):740-6. doi: [10.1016/j.ceb.2003.10.006](#)
77. Okamoto I, Pirker C, Bilban M, Berger W, Losert D, Marosi C, et al. Seven novel and stable translocations associated with oncogenic gene expression in malignant melanoma. *Neoplasia* 2005;7(4):303-11. doi: [10.1593/neo.04514](#)
78. Li D, Xia L, Chen M, Lin C, Wu H, Zhang Y, et al. miR-133b, a particular member of myomiRs, coming into playing its unique pathological role in human cancer. *Oncotarget* 2017;8(30):50193-208. doi: [10.18632/oncotarget.16745](#)
79. Wang X, Bu J, Liu X, Wang W, Mai W, Lv B, et al. miR-133b suppresses metastasis by targeting HOXA9 in human colorectal cancer. *Oncotarget* 2017;8(38):63935-48. doi: [10.18632/oncotarget.19212](#)
80. Baharudin R, Rus Bakaruraini NQ, Ismail I, Lee LH, Ab Mutalib NS. MicroRNA methylome signature and their functional roles in colorectal cancer diagnosis, prognosis, and chemoresistance. *Int J Mol Sci* 2022;23(13):7281. doi: [10.3390/ijms23137281](#)
81. Mercer TR, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nat Rev Genet* 2009;10(3):155-9. doi: [10.1038/nrg2521](#)
82. Jalaie A, Asadi MR, Sabaie H, Dehghani H, Gharepouran J, Hussen BM, et al. Long non-coding RNAs, novel offenders or guardians in multiple sclerosis: a scoping review. *Front Immunol* 2021;12:774002. doi: [10.3389/fimmu.2021.774002](#)
83. Li J, Li Z, Leng K, Xu Y, Ji D, Huang L, et al. ZEB1-AS1: a crucial cancer-related long non-coding RNA. *Cell Prolif* 2018;51(1):e12423. doi: [10.1111/cpr.12423](#)
84. Shi K, Yang S, Chen C, Shao B, Guo Y, Wu X, et al. RNA methylation-mediated LINC01559 suppresses colorectal cancer progression by regulating the miR-106b-5p/PTEN axis.

- Int J Biol Sci* 2022;18(7):3048-65. doi: [10.7150/ijbs.70630](https://doi.org/10.7150/ijbs.70630)
85. Pedersen SK, Mitchell SM, Graham LD, McEvoy A, Thomas ML, Baker RT, et al. CAHM, a long non-coding RNA gene hypermethylated in colorectal neoplasia. *Epigenetics* 2014;9(8):1071-82. doi: [10.4161/epi.29046](https://doi.org/10.4161/epi.29046)
 86. Sun J, Ding C, Yang Z, Liu T, Zhang X, Zhao C, et al. The long non-coding RNA TUG1 indicates a poor prognosis for colorectal cancer and promotes metastasis by affecting epithelial-mesenchymal transition. *J Transl Med* 2016;14:42. doi: [10.1186/s12967-016-0786-z](https://doi.org/10.1186/s12967-016-0786-z)
 87. Lv L, He L, Chen S, Yu Y, Che G, Tao X, et al. Long non-coding RNA LINC00114 facilitates colorectal cancer development through EZH2/DNMT1-induced miR-133b suppression. *Front Oncol* 2019;9:1383. doi: [10.3389/fonc.2019.01383](https://doi.org/10.3389/fonc.2019.01383)
 88. Li CF, Li YC, Wang Y, Sun LB. The effect of lncRNA H19/miR-194-5p axis on the epithelial-mesenchymal transition of colorectal adenocarcinoma. *Cell Physiol Biochem* 2018;50(1):196-213. doi: [10.1159/000493968](https://doi.org/10.1159/000493968)
 89. Zhang H, Lu Y, Wu J, Feng J. LINC00460 hypomethylation promotes metastasis in colorectal carcinoma. *Front Genet* 2019;10:880. doi: [10.3389/fgene.2019.00880](https://doi.org/10.3389/fgene.2019.00880)
 90. Wu Y, Yang X, Chen Z, Tian L, Jiang G, Chen F, et al. m6A-induced lncRNA RP11 triggers the dissemination of colorectal cancer cells via upregulation of Zeb1. *Mol Cancer* 2019;18(1):87. doi: [10.1186/s12943-019-1014-2](https://doi.org/10.1186/s12943-019-1014-2)
 91. Wu X, Li R, Song Q, Zhang C, Jia R, Han Z, et al. JMJD2C promotes colorectal cancer metastasis via regulating histone methylation of MALAT1 promoter and enhancing β -catenin signaling pathway. *J Exp Clin Cancer Res* 2019;38(1):435. doi: [10.1186/s13046-019-1439-x](https://doi.org/10.1186/s13046-019-1439-x)
 92. Guo T, Liu DF, Peng SH, Xu AM. ALKBH5 promotes colon cancer progression by decreasing methylation of the lncRNA NEAT1. *Am J Transl Res* 2020;12(8):4542-9.
 93. Xu L, Huan L, Guo T, Wu Y, Liu Y, Wang Q, et al. lncRNA SNHG11 facilitates tumor metastasis by interacting with and stabilizing HIF-1 α . *Oncogene* 2020;39(46):7005-18. doi: [10.1038/s41388-020-01512-8](https://doi.org/10.1038/s41388-020-01512-8)
 94. Shigeyasu K, Toden S, Ozawa T, Matsuyama T, Nagasaka T, Ishikawa T, et al. The PVT1 lncRNA is a novel epigenetic enhancer of MYC, and a promising risk-stratification biomarker in colorectal cancer. *Mol Cancer* 2020;19(1):155. doi: [10.1186/s12943-020-01277-4](https://doi.org/10.1186/s12943-020-01277-4)
 95. Galamb O, Kalmár A, Sebestyén A, Dankó T, Kriston C, Fúri I, et al. Promoter hypomethylation and increased expression of the long non-coding RNA LINC00152 support colorectal carcinogenesis. *Pathol Oncol Res* 2020;26(4):2209-23. doi: [10.1007/s12253-020-00800-8](https://doi.org/10.1007/s12253-020-00800-8)
 96. Song P, Li Y, Wang F, Pu L, Bao L, Gao H, et al. Genome-wide screening for differentially methylated long noncoding RNAs identifies LIFR-AS1 as an epigenetically regulated lncRNA that inhibits the progression of colorectal cancer. *Clin Epigenetics* 2022;14(1):138. doi: [10.1186/s13148-022-01361-0](https://doi.org/10.1186/s13148-022-01361-0)
 97. Stelzer G, Rosen N, Plaschkes I, Zimmerman S, Twik M, Fishilevich S, et al. The GeneCards suite: from gene data mining to disease genome sequence analyses. *Curr Protoc Bioinformatics* 2016;54(1):1-30. doi: [10.1002/cpbi.5](https://doi.org/10.1002/cpbi.5)
 98. Krishnamachary B, Berg-Dixon S, Kelly B, Agani F, Feldser D, Ferreira G, et al. Regulation of colon carcinoma cell invasion by hypoxia-inducible factor 1. *Cancer Res* 2003;63(5):1138-43.
 99. Liu K, Yao H, Wen Y, Zhao H, Zhou N, Lei S, et al. Functional role of a long non-coding RNA LIFR-AS1/miR-29a/TNFAIP3 axis in colorectal cancer resistance to photodynamic therapy. *Biochim Biophys Acta Mol Basis Dis* 2018;1864(9 Pt B):2871-80. doi: [10.1016/j.bbdis.2018.05.020](https://doi.org/10.1016/j.bbdis.2018.05.020)
 100. Bao-Caamano A, Costa-Fraga N, Cayrefourcq L, Rodriguez-Casanova A, Muñelo-Romay L, López-López R, et al. Epigenomic reprogramming of therapy-resistant circulating tumor cells in colon cancer. *Front Cell Dev Biol* 2023;11:1291179. doi: [10.3389/fcell.2023.1291179](https://doi.org/10.3389/fcell.2023.1291179)
 101. Guo L, Lee YT, Zhou Y, Huang Y. Targeting epigenetic regulatory machinery to overcome cancer therapy resistance. *Semin Cancer Biol* 2022;83:487-502. doi: [10.1016/j.semcancer.2020.12.022](https://doi.org/10.1016/j.semcancer.2020.12.022)
 102. Luo M, Yang X, Chen HN, Nice EC, Huang C. Drug resistance in colorectal cancer: an epigenetic overview. *Biochim Biophys Acta Rev Cancer* 2021;1876(2):188623. doi: [10.1016/j.bbcan.2021.188623](https://doi.org/10.1016/j.bbcan.2021.188623)
 103. Pan S, Deng Y, Fu J, Zhang Y, Zhang Z, Qin X. N6-methyladenosine upregulates miR-181d-5p in exosomes derived from cancer-associated fibroblasts to inhibit 5-FU sensitivity by targeting NCALD in colorectal cancer. *Int J Oncol* 2022;60(2):14. doi: [10.3892/ijo.2022.5304](https://doi.org/10.3892/ijo.2022.5304)
 104. Lai J, Zhou Z, Hu K, Yu H, Su X, Niu X, et al. N6-methyladenosine methylation analysis of long noncoding RNAs and mRNAs in 5-FU-resistant colon cancer cells. *Epigenetics* 2024;19(1):2298058. doi: [10.1080/15592294.2023.2298058](https://doi.org/10.1080/15592294.2023.2298058)
 105. Takahashi M, Cuatrecasas M, Balaguer F, Hur K, Toiyama Y, Castells A, et al. The clinical significance of miR-148a as a predictive biomarker in patients with advanced colorectal cancer. *PLoS One* 2012;7(10):e46684. doi: [10.1371/journal.pone.0046684](https://doi.org/10.1371/journal.pone.0046684)
 106. Zhang Y, Yu J, Liu H, Ma W, Yan L, Wang J, et al. Novel epigenetic CREB-miR-630 signaling axis regulates radiosensitivity in colorectal cancer. *PLoS One* 2015;10(8):e0133870. doi: [10.1371/journal.pone.0133870](https://doi.org/10.1371/journal.pone.0133870)
 107. Ma Y, Yang Y, Wang F, Moyer MP, Wei Q, Zhang P, et al. Long non-coding RNA CCLAL regulates colorectal cancer progression by activating Wnt/ β -catenin signalling pathway via suppression of activator protein 2 α . *Cut* 2016;65(9):1494-504. doi: [10.1136/gutjnl-2014-308392](https://doi.org/10.1136/gutjnl-2014-308392)
 108. Lu Y, Zhao X, Liu Q, Li C, Graves-Deal R, Cao Z, et al. lncRNA MIR100HG-derived miR-100 and miR-125b mediate cetuximab resistance via Wnt/ β -catenin signaling. *Nat Med* 2017;23(11):1331-41. doi: [10.1038/nm.4424](https://doi.org/10.1038/nm.4424)
 109. Shi L, Li X, Wu Z, Li X, Nie J, Guo M, et al. DNA methylation-mediated repression of miR-181a/135a/302c expression promotes the microsatellite-unstable colorectal cancer development and 5-FU resistance via targeting PLAG1. *J Genet Genomics* 2018;45(4):205-14. doi: [10.1016/j.jgg.2018.04.003](https://doi.org/10.1016/j.jgg.2018.04.003)
 110. Yu W, Shi Q, Wu C, Shen X, Chen L, Xu J. Promoter hypermethylation influences the suppressive role of long non-coding RNA MEG3 in the development of multiple myeloma. *Exp Ther Med* 2020;20(1):637-45. doi: [10.3892/etm.2020.8723](https://doi.org/10.3892/etm.2020.8723)
 111. Wang H, Li H, Zhang L, Yang D. Overexpression of MEG3 sensitizes colorectal cancer cells to oxaliplatin through regulation of miR-141/PDCD4 axis. *Biomed Pharmacother* 2018;106:1607-15. doi: [10.1016/j.biopha.2018.07.131](https://doi.org/10.1016/j.biopha.2018.07.131)
 112. Shan S, Lu Y, Zhang X, Shi J, Li H, Li Z. Inhibitory effect of bound polyphenol from foxtail millet bran on miR-149 methylation increases the chemosensitivity of human colorectal cancer HCT-8/Fu cells. *Mol Cell Biochem* 2021;476(2):513-23. doi: [10.1007/s11010-020-03906-4](https://doi.org/10.1007/s11010-020-03906-4)
 113. Guttman M, Donaghey J, Carey BW, Garber M, Grenier JK, Munson G, et al. lncRNAs act in the circuitry controlling pluripotency and differentiation. *Nature* 2011;477(7364):295-300. doi: [10.1038/nature10398](https://doi.org/10.1038/nature10398)
 114. Schetter AJ, Leung SY, Sohn JJ, Zanetti KA, Bowman ED, Yanaihara N, et al. MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. *JAMA* 2008;299(4):425-36. doi: [10.1001/jama.299.4.425](https://doi.org/10.1001/jama.299.4.425)
 115. Nishida N, Mimori K, Fabbri M, Yokobori T, Sudo T, Tanaka F, et al. MicroRNA-125a-5p is an independent prognostic

- factor in gastric cancer and inhibits the proliferation of human gastric cancer cells in combination with trastuzumab. *Clin Cancer Res* 2011;17(9):2725-33. doi: [10.1158/1078-0432.Ccr-10-2132](https://doi.org/10.1158/1078-0432.Ccr-10-2132)
116. Lu JH, Zuo ZX, Wang W, Zhao Q, Qiu MZ, Luo HY, et al. A two-microRNA-based signature predicts first-line chemotherapy outcomes in advanced colorectal cancer patients. *Cell Death Discov* 2018;4:116. doi: [10.1038/s41420-018-0133-7](https://doi.org/10.1038/s41420-018-0133-7)
 117. Rokavec M, Li H, Jiang L, Hermeking H. The p53/miR-34 axis in development and disease. *J Mol Cell Biol* 2014;6(3):214-30. doi: [10.1093/jmcb/mju003](https://doi.org/10.1093/jmcb/mju003)
 118. Fu J, Imani S, Wu MY, Wu RC. MicroRNA-34 family in cancers: role, mechanism, and therapeutic potential. *Cancers (Basel)* 2023;15(19):4723. doi: [10.3390/cancers15194723](https://doi.org/10.3390/cancers15194723)
 119. Krajewska JB, Fichna J, Mosińska P. One step ahead: miRNA-34 in colon cancer-future diagnostic and therapeutic tool? *Crit Rev Oncol Hematol* 2018;132:1-8. doi: [10.1016/j.critrevonc.2018.09.006](https://doi.org/10.1016/j.critrevonc.2018.09.006)
 120. Sabaie H, Amirinejad N, Asadi MR, Jalaie A, Daneshmandpour Y, Rezaei O, et al. Molecular insight into the therapeutic potential of long non-coding RNA-associated competing endogenous RNA axes in Alzheimer's disease: a systematic scoping review. *Front Aging Neurosci* 2021;13:742242. doi: [10.3389/fnagi.2021.742242](https://doi.org/10.3389/fnagi.2021.742242)
 121. Chen H, Xu Z, Liu D. Small non-coding RNA and colorectal cancer. *J Cell Mol Med* 2019;23(5):3050-7. doi: [10.1111/jcmm.14209](https://doi.org/10.1111/jcmm.14209)
 122. Ferreira HJ, Heyn H, Moutinho C, Esteller M. CpG island hypermethylation-associated silencing of small nucleolar RNAs in human cancer. *RNA Biol* 2012;9(6):881-90. doi: [10.4161/rna.19353](https://doi.org/10.4161/rna.19353)
 123. Tang Q, Li L, Wang Y, Wu P, Hou X, Ouyang J, et al. RNA modifications in cancer. *Br J Cancer* 2023;129(2):204-21. doi: [10.1038/s41416-023-02275-1](https://doi.org/10.1038/s41416-023-02275-1)
 124. Zhu DH, Su KK, Ou-Yang XX, Zhang YH, Yu XP, Li ZH, et al. Mechanisms and clinical landscape of N6-methyladenosine (m6A) RNA modification in gastrointestinal tract cancers. *Mol Cell Biochem* 2024;479(7):1553-70. doi: [10.1007/s11010-024-05040-x](https://doi.org/10.1007/s11010-024-05040-x)
 125. Zhang X, Su H, Chen H, Li Q, Liu X, Zhang L, et al. RNA modifications in gastrointestinal cancer: current status and future perspectives. *Biomedicines* 2022;10(8):1918. doi: [10.3390/biomedicines10081918](https://doi.org/10.3390/biomedicines10081918)
 126. Azizidoost S, Ghaedrahmati F, Anbiyaei O, Ahmad Ali R, Cheraghzadeh M, Farzaneh M. Emerging roles for lncRNA-NEAT1 in colorectal cancer. *Cancer Cell Int* 2022;22(1):209. doi: [10.1186/s12935-022-02627-6](https://doi.org/10.1186/s12935-022-02627-6)